

Na+/K+ ATPase Microplate Assay Kit

Catalog # AS0019

Detection and Quantification of Na+/K+ ATPase Activity in Urine,
Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and
Other biological fluids Samples.

This instruction must be read in its entirety before using this product.

For research use only, Not for use in diagnostic procedures.

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I. INTRODUCTION

Na⁺/K⁺ ATPase is widely distributed in plants, animals, microbes and cells,can catalyze the hydrolysis of ATP, ADP and inorganic phosphate.

Na+/K+ ATPase catalyze the decomposition of ATP into ADP and free phosphate ion. These enzymes play key roles in transport, signal transduction, protein biosynthesis and cell differentiation. At the end of the reaction period, the dye reagent forms a color with released phosphate ion, which is measured on a plate reader 660 nm.



II.KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Substrate	Powder x 1	-20 °C
Activator	Powder x 1	4 °C
Inhibitor	Powder x 1	4 °C
Dye Reagent I	Powder x 1	4 °C
Dye Reagent II	Powder x 1	4 °C
Dye Reagent III	20 ml x 1	4 °C
Stop Solution	4 ml x 1	RT
Standard (5µmol/ml)	1 ml x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Substrate: add 17 ml distilled water to dissolve before use, store at 4 °C.

Activator: add 1 ml distilled water to dissolve before use, store at 4 °C. (If it is difficult to dissolve, please heat up.)

Inhibitor:add 1 ml distilled water to dissolve before use, store at 4 °C. (If it is difficult to dissolve, please heat up.)

Dye Reagent: add 10 ml Dye Reagent III into Dye Reagent I and 2 ml Dye Reagent III into Dye Reagent II respectivelyto dissolve. Transfer allDye Reagent IIInto Dye Reagent III, mix, then transfer all Dye Reagent I into Dye Reagent III (Must follow this step). The mixed Dye Reagent may store at 4 °C for 2-3 days.

*Note:It should be yellow. If colorless, the solution is failure. If blue, the solutionispolluted. This solution should be prepared before use. It is best to used is possible plastic containers to prepare the solution in order to prevent phosphorus pollution.



III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 660 nm
- 2. Distilled water
- 3. Pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Ice
- 7. Centrifuge

IV. SAMPLE PREPARATION

1.For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 mlAssay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s,repeat 30 times); centrifuged at 8000g 4°C for 10minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weighout 0.1 g tissue, homogenize with 1 mlAssay buffer on ice, centrifuged at 8000g 4°C for 10minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For red blood cell samples

Add heparin into the blood, centrifuged at 2000g 4 °C for 5 minutes. Discard the plasma and white blood cells. Wash the red blood cells with PBS for 3 times, discard the supernatant after centrifugation each time. Add 0.9 ml Assay buffer into 0.1 ml red blood cells, mix well, and wait for 15 minutes at room temperature.



V. ASSAY PROCEDURE

Add following reagents into the microcentrifuge tubes:

Reagent	Sample	Control	Standard	Blank		
Substrate	170μΙ	170μΙ				
Sample	20μΙ	20μΙ				
Inhibitor		10μΙ				
Activator	10μΙ					
Mix, put it in the oven,37°Cfor 30 minutes.						
Stop Solution	40 μΙ	40 μΙ				
Mix, centrifuged at 10000g, room temperature for5 minutes. Add following						
reagents intothe microplate:						
Standard			20 μΙ			
Distilled water				20 μΙ		
Supernatant	20 μΙ	20 μΙ				
Dye Reagent	180 μΙ	180 μΙ	180 μΙ	180 μΙ		
Mix, room temperaturefor 30 minutes, record absorbance measured at 660nm.						

Note: It is best to use disposable plastic tube to avoid phosphorus pollution.



VI. CALCULATION

Unit Definition:One unit of Na $^+$ /K $^+$ ATPaseactivityis defined as the enzyme generates 1 μ mol of PO $_4$ 3-per hour.

1. According to the protein concentration of sample

$$\begin{split} \text{Na}^+/\text{K}^+ \text{ ATPase (U/mg) = } & (C_{\text{Standard}} \times V_{\text{Total}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) \text{ / } (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \text{ / } \\ & (V_{\text{Sample}} \times C_{\text{Protein}}) \text{ / } T \\ & = 120 \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Control}}) \text{ / } (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \text{ / } C_{\text{Protein}} \end{split}$$

2. According to the weight of sample

Na*/K* ATPase (U/g) =(
$$C_{Standard} \times V_{Total}$$
)×(OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / (W $\times V_{Sample}$ / V_{Assay}) / T = 120×(OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / W

3. According to the concentration of cell or bacteria

$$Na^{+}/K^{+} \text{ ATPase (U/10^{4}) = (C_{Standard} \times V_{Total}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / }$$

$$(N \times V_{Sample} / V_{Assay}) / T$$

$$= 120 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / N$$

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

C_{Standard}: the concentration of Standard, 5µmol/ml;

V_{Total}: the total volume of the enzymatic reaction, 0.24 ml;

V_{Sample}: the volume of sample, 0.02 ml;

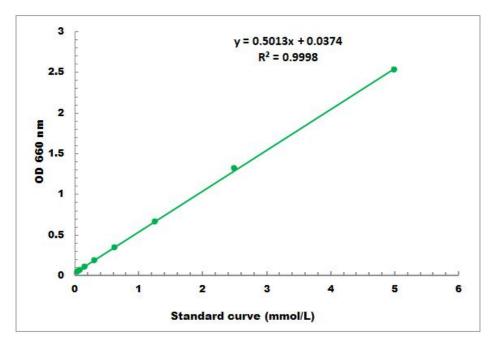
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 0.5 h.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 μmol/ml - 5μmol/ml

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.sabbiotech.cn or contact us at techcn@signalwayantibody.com

IX. NOTES