

Chitinase Microplate Assay Kit

Catalog # AS0088

Detection and Quantification of Chitinase Activity in Tissue extracts,
Cell lysate, Cell culture media and Other biological fluids Samples.

This instruction must be read in its entirety before using this product.

For research use only, Not for use in diagnostic procedures.

Contact information:

Tel:+1 (301) 446-2499 Fax:+1 (301) 446-2413

Email:techcn@signalwayantibody.com Web:www.sabbiotech.com



| I. INTRODUCTION | 2 |
|--|---|
| II. KIT COMPONENTS | 3 |
| III. MATERIALS REQUIRED BUT NOT PROVIDED | |
| VI. SAMPLE PREPARATION | 4 |
| V. ASSAY PROCEDURE | 5 |
| VI. CALCULATION | 6 |
| VII. TYPICAL DATA | 7 |
| VIII. TECHNICAL SUPPORT | 7 |
| IX NOTES | 7 |



I. INTRODUCTION

Chitinases (EC 3.2.1.14) are hydrolytic enzymes that break down glycosidic bonds in chitin. Chitinase catalyzes the hydrolytic cleavage of thebeta-1→4-glycoside bond present in biopolymers of N-acetylglucosamine, primarily in chitin. Chitinases are widely distributed in living organisms and are found in fungi, bacteria, parasites, plants, and animals. They are classified in families based on amino acid sequence similarities.

As chitin is a component of the cell walls of fungi and exoskeletal elements of some animals (including worms and arthropods), chitinases are generally found in organisms that either need to reshape their own chitin or dissolve and digest the chitin of fungi or animals.

Chitinases perform different functions in different organisms. In bacteria, they are mainly involved in nutritional processes. In yeast and various fungi, these enzymes participate in morphogenesis. In animals and plants, chitinases primarily play a role in the defense of the organism against pathogen attack.

The assay is initiated with the enzymatic hydrolysis of the chitin by chitinases. The enzyme catalysed reaction productsN-acetylglucosaminereact with PDAB, and can be measured at a colorimetric readout at 585 nm.



II.KIT COMPONENTS

| Component | Volume | Storage | |
|-----------------------|-----------|--------------------|--|
| 96-Well Microplate | 1 plate | | |
| Assay Buffer | 30mlx 4 | 4 °C | |
| Substrate | 10 ml x 1 | 4 °C | |
| Reaction Buffer | 4mlx 1 | 4 °C | |
| Dye Reagent | Powderx 1 | 4 °C, keep in dark | |
| Dye Reagent Diluent | 12 ml x 1 | 4 °C, keep in dark | |
| Standard | Powderx 1 | 4 °C | |
| Plate Adhesive Strips | 3 Strips | | |
| Technical Manual | 1 Manual | | |

Note:

Dye Reagent: add 12 mlDye Reagent Diluent to dissolve before use, store at 4 °C.

Standard: add 1 mldistilled water to dissolve before use, mix; the concentration will be 1 mg/ml, store at 4 °C.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 585 nm
- 2. Distilled water
- 3. Pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 mlAssay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s,repeat 30 times); centrifuged at 12,000g 4°C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2.For tissue samples

Weighout 0.1 g tissue, homogenize with 1 mlAssay buffer on ice, centrifuged at 12,000g 4°C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For cell culture media

Detect directly.



V. ASSAY PROCEDURE

Dye Reagent

Add following reagents into the microcentrifuge tube:

120 μΙ

| Reagent | Sample | Control | Standard | Blank | | | |
|--|--------|---------|----------|-------|--|--|--|
| Sample | 80 μΙ | | | | | | |
| Assay Buffer | | 80 μΙ | | | | | |
| Substrate | 80 μΙ | 80 μΙ | | | | | |
| Mix, put it in the oven, 37°C for 1 hour. Centrifuged at 5000g, 4°C for 10minutes, | | | | | | | |
| add80 μlsupernatant into the new microcentrifuge tube. | | | | | | | |
| Supernatant | 80 μΙ | 80 μΙ | | | | | |
| Standard | | | 80 μΙ | | | | |
| Distilled water | | | | 80 μΙ | | | |
| Reaction Buffer | 40 μΙ | 40 μΙ | 40 μΙ | 40 μΙ | | | |
| Mix, put it in the boiling water for 7 minutes. Centrifuged at 5000g for | | | | | | | |
| 2minutesaddthesupernatant into the microplate. | | | | | | | |
| Supernatant | 80 μΙ | 80 μΙ | 80 μΙ | 80 μΙ | | | |

Mix, put it in the oven, 37°C for 60 minutes, record absorbance measured at 585 nm.

120 μΙ

120 μΙ

120 μΙ



VI. CALCULATION

Unit Definition:One unit of Chitinase activity is defined as the enzymegenerates $1\mu g$ of N-acetylglucosamine per hour at $37^{\circ}C$.

1. According to the protein concentration of sample

Chitinase (U/mg) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} / C_{Protein} / T \times 2$$

2. According to the weight of sample

Chitinase (U/g) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times W / V_{Assay}) / T \times 2$$
= 2000×(OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / W

3. According to the quantity of cells or bacteria

Chitinase (U/10⁴)=(
$$C_{Standard} \times V_{Standard}$$
) × (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / ($V_{Sample} \times N / V_{Assay}$) / T × 2 = 2000×(OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /N

C_{Protein}: the protein concentration, mg/ml;

 $C_{Standard}$: the concentration of Standard, 1 mg/ml = 1000 μ g/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria, N ×10⁴;

V_{Standard}: the volume of standard, 0.08 ml;

V_{Sample}: the volume of sample, 0.08 ml;

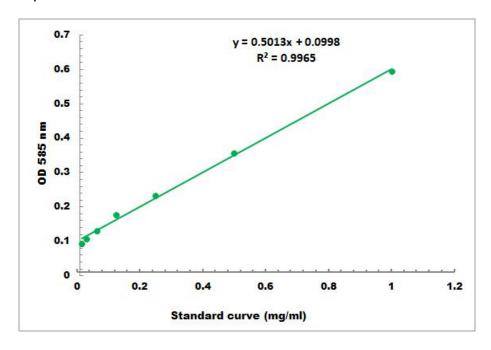
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 1 h.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01mg/ml - 1mg/ml

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.sabbiotech.cn or contact us at techcn@signalwayantibody.com

IX. NOTES